Felix Grünberger¹, Zubeir El Ahmad¹, Winfried Hausner¹, Sébastien Ferreira-Cerca², Dina Grohmann¹ ¹ Institute of Microbiology and Archaea Centre, Regensburg, Germany | ² Biochemistry, Regensburg, Germany | Contact: **felix.gruenberger@ur.de**





Expanding the transcriptomic toolbox in prokaryotes by Nanopore sequencing of RNA and cDNA molecules

Nanopore sequencing is a 3rd generation single-molecule method that allows sequencing of full-length transcripts.

Here, we present an experimental and bioinformatic workflow for ONT RNA-seq in the bacterial model organism Escherichia coli (i), and show how we used the technolgy to analyze the stage-dependent installation of rRNA modifications (1) and transcription termination heterogeneity in Archaea (2).



Literature

 $\mathbf{\hat{U}}$ This study

Application 1: Stage-dependent installation of rRNA modifications

Ribosome biogenesis in Archaea starts with a polycistronic pre-rRNA, that is processed via the coordinated and defined order of **ribonucleases** action, **RNA folding** and **RNA base modifications**.



Using direct RNA sequencing, we followed the maturation pathway K in Pyrococcus furiosus and analyzed the stage-dependent installation of selected modifications in helix 45 of the 16S rRNA.

RNA modifications can lead to electric current signals varying from the expected theoretical distribution **L** and consequently may increase the rate of **basecalling errors M**.



dependent N⁴-cytidine

already installed earlier.

24日1月

•

acetylation (N^4-Ac) is

Application 2: Transcription termination heterogeneity

aCPSF1

RNAP

The efficiency of transcription termination in Archaea presumably depends on the cooperative action of the termination factor **aCPSF1** that specificially recognizes and cleaves after **poly**uridine tracts, which are the intrinsic termination signal in Archaea.



Here, we demonstrate the applicability of a

 \triangleright Single-molecule sequencing captures termination heterogeneity



Termination efficiency is poly(U) and



modified and improved polyA-independent PCR-cDNA ONT protocol, to accurately map transcript **3' ends** in *Pyrococcus furiosus* at



▷Improved 3'end ▷ Improved proportion of full-length reads accuracy (Comparison to short-(in %) read ends)



temperature-dependent

